Azacytidine for Concurrent Acute Monocytic Leukemia and Multiple Myeloma in an Elderly Patient

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To the editor,

Simultaneous occurrence of MM and AML is a rare entity.¹,² Herein, we report an elderly patient with simultaneous MM and AML, successfully treated with azacytidine.

A seventy-one year old male patient was referred with fatigue, immune-paresis and IgG-Lambda monoclonal gammopathy. Physical examination was unremarkable. Complete blood counts were hemoglobin: 13 g/dL (12-16), platelets: 175x10⁹/L (150-450), leukocytes: 3.3×10⁹/L (3.91-8.77), neutrophils: 1.8×10⁹/L (2.06-7.02), monocytes: 0.5×10⁹/L (0.3-0.8). Serum calcium and creatinine levels were normal. Several lytic lesions were detected in cervical vertebrae by magnetic resonance imaging. Bone marrow samples showed CD38, CD138, CD56 and lambda positive, CD19 negative, 10-20% monoclonal plasma cells. Karyotyping revealed Y chromosome loss. Fluorescent in situ hybridization (FISH) was negative for common, and/or prognostically significant chromosomal aberrations. Patient was diagnosed with symptomatic MM and bortezomib-cyclophosphamide-dexamethasone (VCD) regimen was started. He was admitted with pneumosepsis and pancytopenia at the end of first cycle. During follow-up, leukocytosis gradually developed within ten days (leukocytes: 35.6x10⁹/L, neutrophils: 9.7x10⁹/L, lymphocytes: 3.2x10⁹/L, monocytes: 0.5x10⁹/L). Monocytes and monoblasts were markedly increased in blood smear. In a repeat bone marrow aspiration, CD34, CD11b, CD11c positive; CD34, CD117, MPO, CD13, CD14 negative, 86% monocytic infiltration was observed; concurrent monoclonal plasma cells were undetectable. Karyotyping was 46,XY and both MM and AML-FISH panel were negative. FLT3-ITD PCR was negative, nucleophosmin (NPM1) PCR was positive for type A mutation.

Figure 1. Monoclonal plasma cell infiltration, along with myeloid maturation arrest a lesser monocytic component was detected (H&E, x200)
When the first bone marrow samples were reexamined, myeloid maturation arrest and a lesser monocytic component was detected; along with monoclonal plasma cells (Figure 1).

Patient was diagnosed as AML and MM. Azacytidine monotherapy (75 mg/m²/day, 7 days, 28-day-cycle) was started. Both MM and AML partially responded (3% clonal plasma cells and 9% monocytic population) to treatment at the end of 4th cycle with complete hematological recovery (Figures 2, 3).

Patient was clinically stable for 15 cycles. After 15 cycles, he was admitted with pneumosepsis and pancytopenia. Bone marrow samples showed both 22% monoclonal plasma cells and 35% monocytes. MM and AML had lost response to therapy, and patient succumbed to septic shock.

It is unclear how AML and MM occur concurrently; several genetic defects were associated with this rare occurrence. Although our patient was diagnosed with AML after the initial cycle of VCD, we classified the patient as concurrent AML and MM, rather than secondary AML to chemotherapy, as we could determine a monocytic component by reviewing previous bone marrow samples. We hypothesize that the proliferation of myeloid clone may have increased by immunosuppression after the initial VCD treatment.

We determined our treatment strategy over AML, since AML would be the determinant of overall survival. Concurrent prognosis is noted to be poor, due to rarity of the disease and no standard treatment. Azacytidine was previously shown to induce apoptosis, upregulate proinflammatory cytokine pathways and cytotoxicity, as well as necrosis in MM cells. Therefore, azacytidine monotherapy was chosen. Azacytidine was effective on both AML and MM clones for 15 months. This regimen could be a reasonable option for future cases with similar diagnosis. Further studies are warranted in order to improve clinicobiological understanding.
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REFERENCES


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Figure 3. Flow cytometric evaluation of bone marrow at (i) the time of MM diagnosis (A), (ii) the time of AML diagnosis (B) and (iii) post-treatment (C). Each row is the continuation of the scatterplot of same markers at different times of treatment. Red color shows MM cells. First row shows CD38 and CD138, second row shows cyIgK and cyIgL, third row shows B2 microglobin and CD56 stains. Percentages are shown below each scatterplot.