Bilateral Sixth Nerve Palsies Possibly Associated with Arsenic Trioxide in Acute Promyelocytic Leukaemia

Pusem PATIR1, Merve OYTUN2, Ahmet A. OZLUK2, Guray SAYDAM1, Ayhan DONMEZ1, Murat TOMBULOGLU3

1 Ege University, Faculty of Medicine, Department of Hematology
2 Ege University, Faculty of Medicine, Department of Internal Medicine, Izmir, TURKEY

Dear Editor,

Acute promyelocytic leukemia (APL) is a unique subtype of acute myeloid leukemia (AML) characterized by specific morphological and cytogenetical aberrations, and a potentially life-threatening coagulopathy.1-3 APL is defined by a specific balanced translocation, t(15;17), resulting in the fusion of PML (promyelocytic leukemia) and RARα (retinoic acid receptor-α) genes. This fusion provides an aberrant, oncogenic protein (PML-RARα) that blocks myeloid differentiation at the promyelocyte stage.4,5 APL is a rare disease (approximately 1200–1500 cases per year in the United States), accounting for approximately 10-15% of all the AML cases.6 All trans retinoic acid (ATRA) and arsenic trioxide (ATO) are the cornerstones of APL therapy, and have dramatically improved outcomes. ATRA in combination with ATO or anthracyclines has been used for the induction treatment of APL, and the rate of complete remission may be as high as 90 % after this treatment.7,8

A 30-year-old female patient was admitted to the hematology department in November of 2014. She was experiencing fatigue, weight loss, hemoptysis and frequent infections. The physical examination revealed normal. Pancytopenia (hemoglobin 11.8 g/dL; leukocytes 1600/mm³; platelet count 36000/mm³) was determined in complete blood count (CBC) and atypical promyelocytic cells with Auer rods were detected on the peripheral blood smear. Bone marrow aspiration and biopsy revealed a hypercellular marrow with 40% promyelocytes containing Auer rods. Flow cytometry was performed to confirm the presence of myeloid blasts (CD34, HLA-DR were negative; CD13, CD33 were positive). Fluorescent in situ hybridization (FISH) and real-time fluorescence quantitative polymerase chain reaction (RT-PCR) analysis were performed in the patient’s bone marrow and confirmed PML-RARα t(15,17).

After a diagnosis of APL was confirmed, ATO was choosen as optional treatment modality and since it was not available in market, an off-label application was delivered to the Ministry of Health (MoH). Since it would take time to have permission from MoH, the induction treatment was started with ATRA as a single agent. From December 1, 2014 to December 14, 2014, she was treated with ATRA 45 mg/m²/day (70 mg/day) for 14 days. On day 15 of the ATRA treatment, she developed a fever, a rapid rise in the leukocyte, and pleuritic chest pain. Firstly, ATRA syndrome was considered and the ATRA was interrupted. She was treated with dexamethasone 20 mg/d for 7 days and then it was discontinued gradually. From December 23, 2014 to December 29, 2014, she was re-treated with ATRA at 45 mg/m²/day (70 mg/day) for 14 days. On day 15 of the ATRA treatment, she developed a fever, a rapid rise in the leukocyte, and pleuritic chest pain. Firstly, ATRA syndrome was considered and the ATRA was interrupted. She was treated with dexamethasone 20 mg/d for 7 days and then it was discontinued gradually. From December 30, 2014 to January 3, 2015, she was treated with ATRA at 45 mg/m²/day (70 mg/day) and ATO at 0.15 mg/kg/day (10 mg/day) for 5 days.
On day 6 of the ATO treatment, hepatotoxicity was observed and improved after the discontinuation of ATRA and ATO and hepato-protective therapy containing N-Acetyl Cysteine. From January 12, 2015 to January 18, 2015, ATRA at 40 mg/day and ATO at 0.075 mg/kg/day (5 mg/day) as a half of a total dose were restarted in combination with methylprednisolone at 40 mg/day for 7 days. From January 19, 2015 to February 15, 2015, ATRA at 40mg/day and ATO at 5mg/day were continued. For the remission control, FISH and RT-PCR analysis were performed on the patient’s bone marrow, and results confirmed complete remission.

On February 16, 2015, the consolidation treatment was started, and she was treated with ATRA at 40 mg/day for 15 days and ATO at 5mg/day for 5 days per week. On the first day of the treatment, she developed sudden blurred vision and diplopia. She was consulted by an ophthalmologist and neurologist. The fundoscopic examination was normal. The neurological examination showed restricted lateral movement of both eyeballs and both apparent abducens nerve palsies. Any other possible findings consistent with peripheral neuropathy were not detected. The neurological and ophthalmological examinations confirmed isolated bilateral 6th nerve palsies, more prominent on the left side, which were at onset, firstly suspected to be caused by isolated leukemic infiltration. The cerebrospinal fluid (CSF) examination and brain magnetic resonance imaging (MRI) were performed in terms of nerve involvement and revealed normal laboratory analysis and imaging. Thus, 6th nerve palsies were considered to be caused by a drug reaction as an adverse event. On February 20, 2015, ATRA and ATO treatment were stopped and then she was monitored. Her complaints completely ceased, and it was decided to replace the consolidation treatment. On March 5, 2015, she was treated with ATRA at 40mg/day for 15 days and idarubicin at 5 mg/m² for 4 days quarterly. The consolidation treatment with ATRA and idarubicin was continued seamlessly; therefore, the adverse ocular reaction was considered to be possibly associated with ATO.

ATO is now a standard drug in the treatment of newly diagnosed or relapsed APL. Treatment with ATO should be monitored carefully due to its potentially severe adverse effect profile. Fluid retention, differentiation syndrome, peripheral sensorial neuropathy, and electrocardiographic abnormalities may be observed as ATO’s side effects during newly diagnosed APL patients’ induction treatment.9 To our knowledge, bilateral sixth nerve palsies have not been previously reported. We detected the chronological relationship, regarding to the onset of ocular symptoms after 40 days of treatment (total dose of ATO was 275 mg), the improvement of ocular symptoms was observed 12 days after discontinuation of ATRA and ATO. Then consolidation treatment was started as ATRA and idarubicin but ocular symptoms were not observed again. So the symptoms were suggestive of the role of ATO, which is known to cause peripheral neurotoxicities. For our case, all other potential reasons of isolated cranial nerve palsy were exclude by CSF analysis and MRI. Peripheral neuropathy is reported in up to 10% of ATO treated patients,10,11 as a neurological side effect. In addition, Thery et al. presented optical neuropathy possibly related to ATO during APL treatment.12 Mechanism of ATO’s neurotoxicity has not discovered yet. In literature, sciatic nerve of rats exposed to ATO were studied and propounded toxicity against neurofilaments bonded with sulfuryl group.13

The blood brain barrier prevents the penetration of heavy metals, including ATO, and protects the central nerve system (CNS). Therefore, CNS side effects are not usually expected during ATO therapy. The entry of the ATO into the CNS however, may occur when the blood brain barrier is disrupted. In a case of meningeal relapse of APL treated with oral-ATO, and the penetration of ATO into the CSF to therapeutically meaningful levels has been observed.14

Unexplained strabismus and diplopia should be evaluated as a potential sign of CNS involvement and initially conventional imaging, and a CSF examination should be performed. If the CNS involvement of APL is not detected, sixth nerve palsy can be considered to be caused by a drug reaction. Herewith, a thorough understanding of the safety and potential side effects of ATO as a therapeutic agent is necessary in order to minimize its toxic complications. In cases with cranial nerve palsy
under the treatment of ATO, after a detailed neurological examination, including MRI and CSF analysis, potential ATO-side effects should be considered.

REFERENCES


Correspondence

Dr. Pusem PATIR
Ege Üniversitesi Tip Fakültesi
Hematoloji Anabilim Dali
35100 Bornova, IZMIR / TURKEY

Telephone: (+90-232) 390 35 40
Fax: (+90-232) 390 35 30
e-mail: pusemp@yahoo.com